

# Sample Preparation

**IDeA** National Resource  
for Proteomics

Workshop for Students and Faculty  
February 2020

# Proteomics Workflow: UAMS Proteomics Core

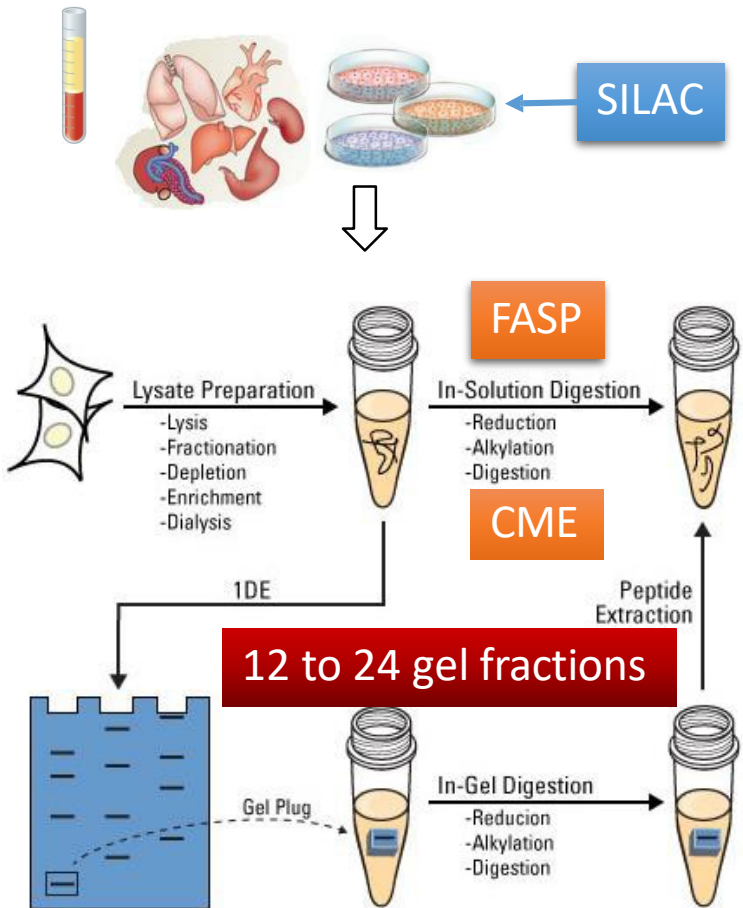
Sample preparation

Enrichment

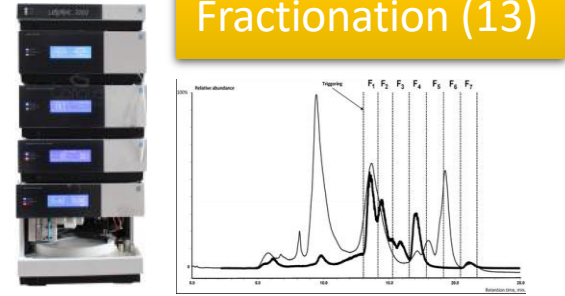
UHPLC-MS/MS

Data analysis

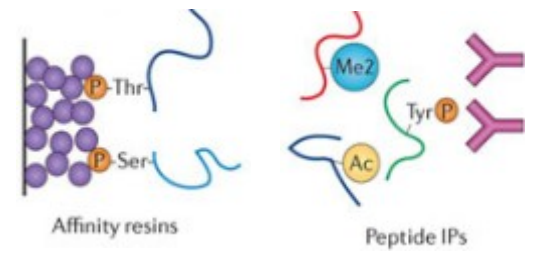
Serum depletion



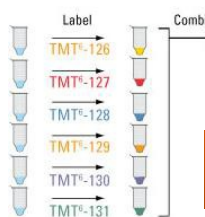
Off-line high pH Fractionation (13)



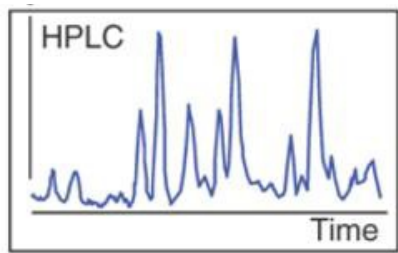
Peptide enrichment



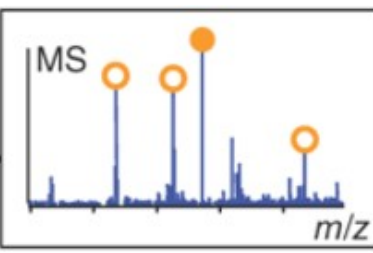
TMT labeling



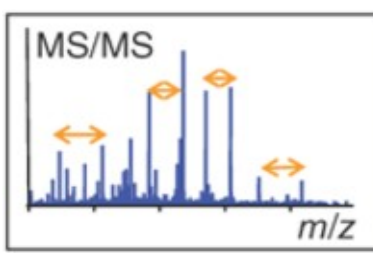
UHPLC separation



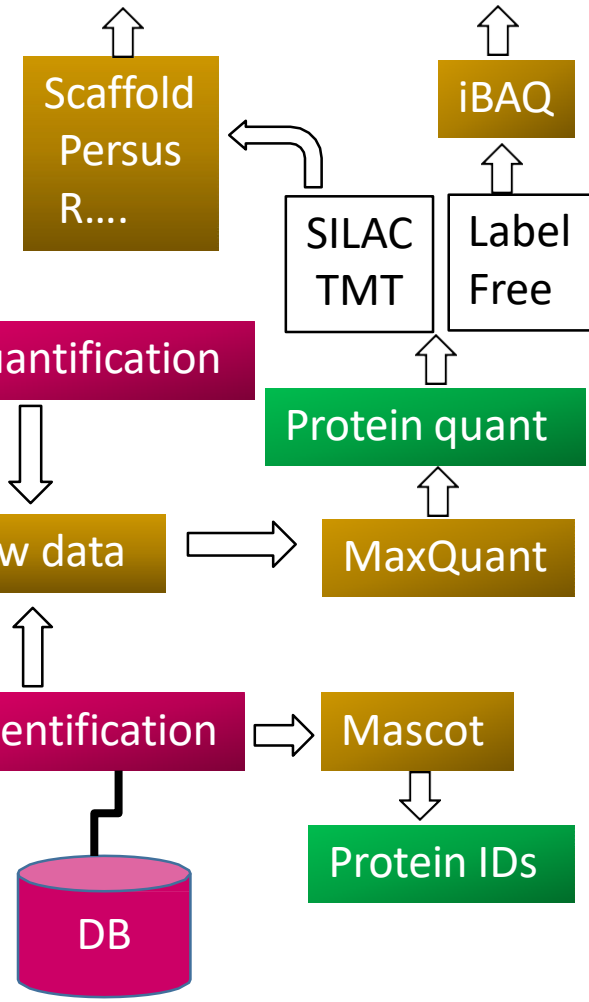
MS1: full spectra



MS/MS: fragments



Comparative proteomics



### Cultured Cells



- cell pellet:  $1-5 \times 10^6$  cells

### Fresh-Frozen Tissue Samples



- 10-100 mg

### FFPE Tissue Samples



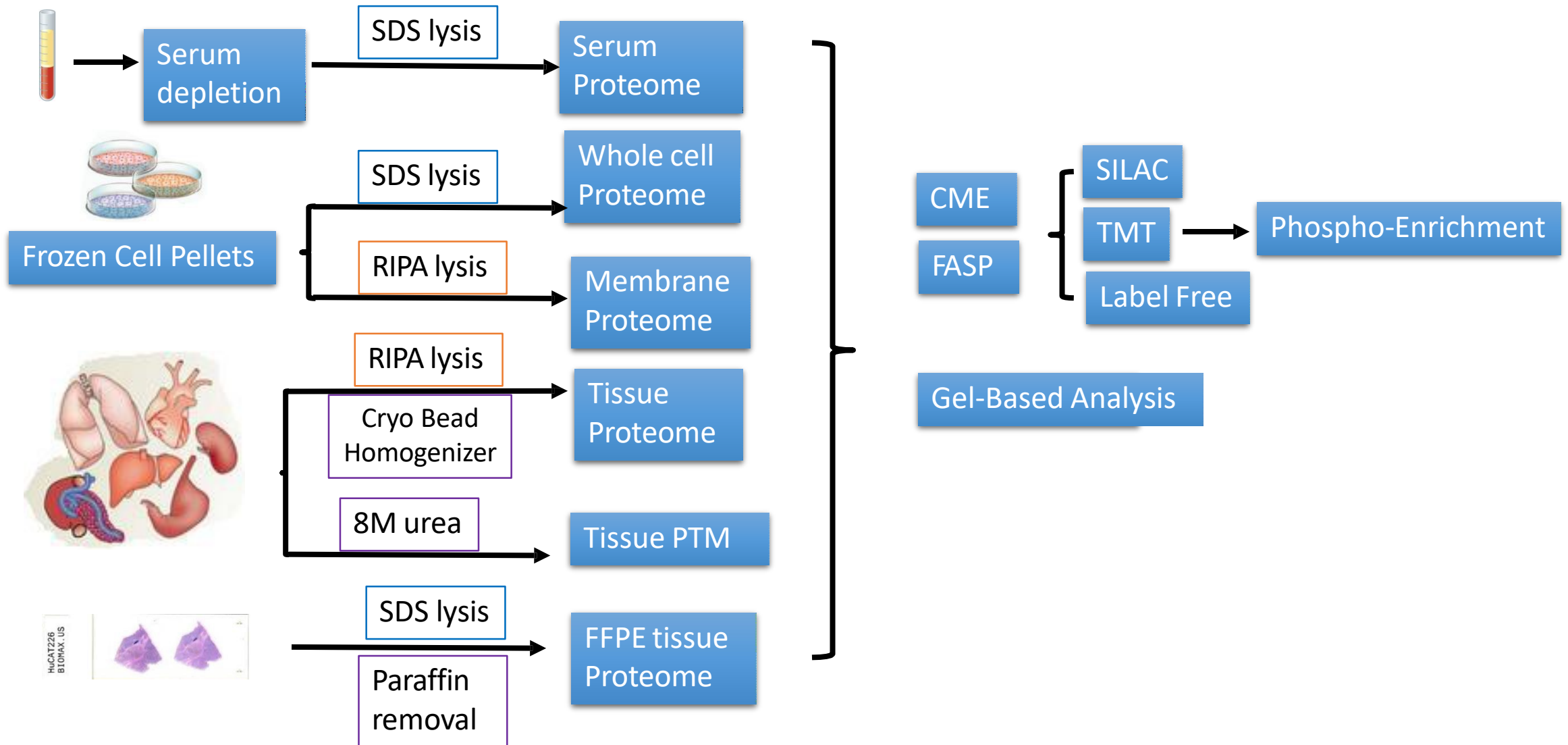
- laser-capture microdissection

### Serum/Plasma Samples



- 10-25 ul

# Decision Tree of Sample Preparation in UAMS Proteomic Core



# In-gel and in-solution digestion

## PROTOCOL

### In-gel digestion for mass spectrometric characterization of proteins and proteomes

Andrej Shevchenko<sup>1,3</sup>, Henrik Tomas<sup>1</sup>, Jan Havliš<sup>1</sup>, Jesper V Olsen<sup>2</sup> & Matthias Mann<sup>2,3</sup>

#### In-gel digestion

- Robustness against impurities
- Fractionate proteins
- Lower peptide recovery
- Lower sequence coverage
- Total solubilization
- stained proteins and assess purity

### Universal sample preparation method for proteome analysis

Jacek R Wiśniewski, Alexandre Zougman, Nagarjuna Nagaraj & Matthias Mann

NATURE METHODS | VOL.6 NO.5 | MAY 2009 | 359

#### In-solution digestion (FASP)

- Can't handle detergents
- Fractionate peptides (high pH UHPLC)
- Higher peptide recovery
- Higher sequence coverage
- Incompletely solubilized



# In-solution digestion (chloroform/methanol extraction)

*Physiol Genomics*. 2007 June 19; 30(1): 89.

## Improved method for the analysis of membrane proteins by mass spectrometry

Shama P. Mirza, Brian D. Halligan, Andrew S. Greene, and Michael Olivier

*MethodsX* 1 (2014) 74–80

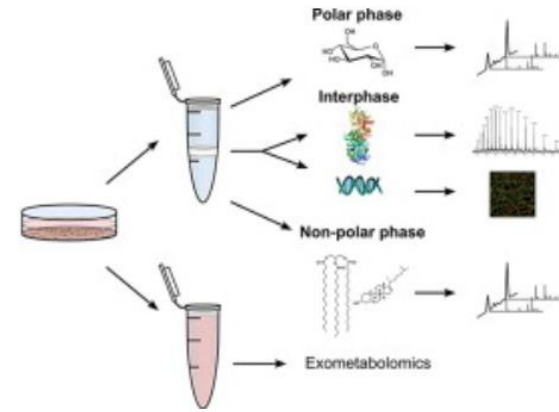
## Simultaneous extraction of proteins and metabolites from cells in culture

Sean C. Sapcariu<sup>a,c,\*</sup>, Tamara Kanashova<sup>b,c</sup>, Daniel Weindl<sup>a</sup>,  
Jenny Ghelfi<sup>a</sup>, Gunnar Dittmar<sup>b,c,\*\*</sup>, Karsten Hiller<sup>a,c</sup>

*J Proteome Res*. 2018 June 01; 17(6): 2226–2236.

## SL-TMT: A Streamlined Protocol for Quantitative (Phospho)proteome Profiling using TMT-SPS-MS3

José Navarrete-Perea<sup>†,#</sup>, Qing Yu<sup>†,#</sup>, Steven P. Gygi<sup>†,\*</sup>, and Joao A. Paulo<sup>†,\*</sup>

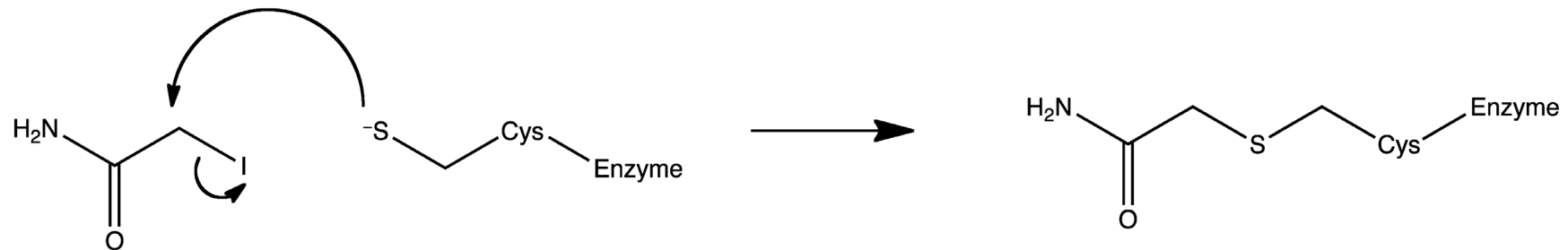


## In-solution digestion (CME)

- Dilipidate proteins
- More detergent tolerance
- Favor membrane proteins
- Simultaneous extract lipids and metabolites

## Reduction and alkylation of disulfide bonds

- Reducing agent: DTT or TCEP
- Alkylating reagent: iodoacetamide



# Protease digestion of proteins for MS analysis

## Trypsin

- most commonly used protease
- cleaves on C-terminal side of basic residues  
Arg and Lys unless next residue is Pro
- active near neutral pH
- inactive at low pH
- sequencing grade trypsin is methylated to protect against self-digestion



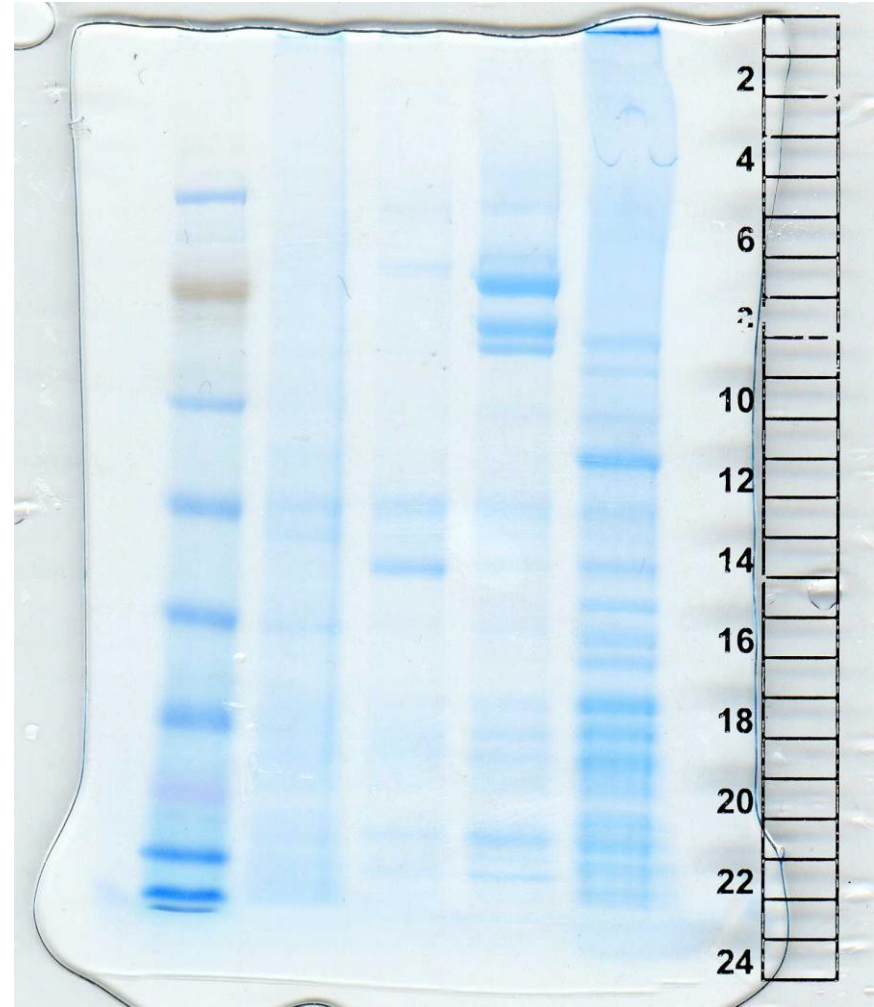
# Protease digestion of proteins for MS analysis

## Commonly used alternate proteases

- AspN – cleaves on N-terminal side of Asp residues
- GluC – cleaves on C-terminal side of Glu residues
- LysC – cleaves on C-terminal side of Lys residues
- ArgC – cleaves on C-terminal side of Arg residues
  
- chymotrypsin – cleaves on C-terminal side of hydrophobic residues
- proteinase K – not sequence specific; used for limited digestion

# Gel-Based Sample Preparation

- Excise gel bands
- Destain gel bands
- Reduction: TCEP
- Alkylation: Iodoacetamide
- Digestion: 100 ng of trypsin
- Quenching: 0.1% formic acid
- LC-MS/MS

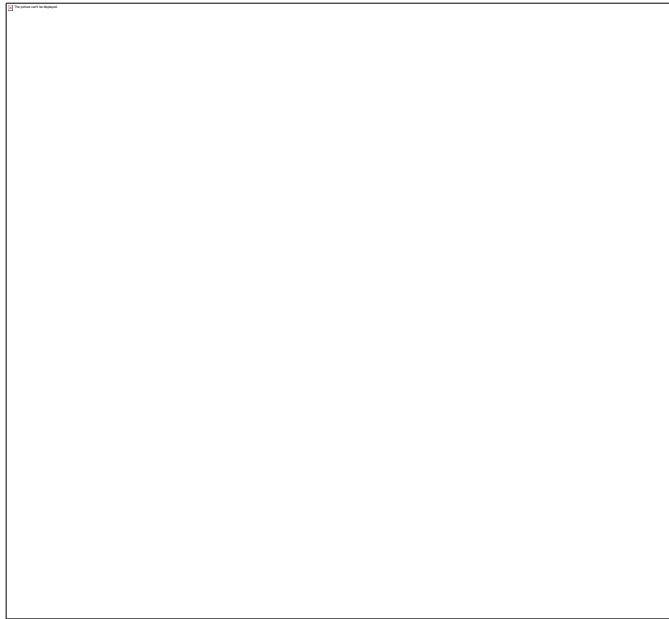


# Gel-Free Sample Cleanup

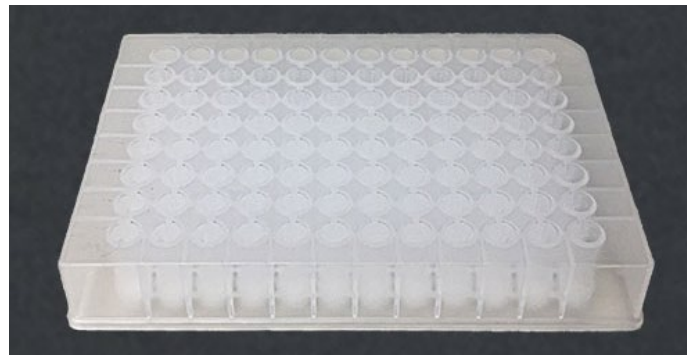
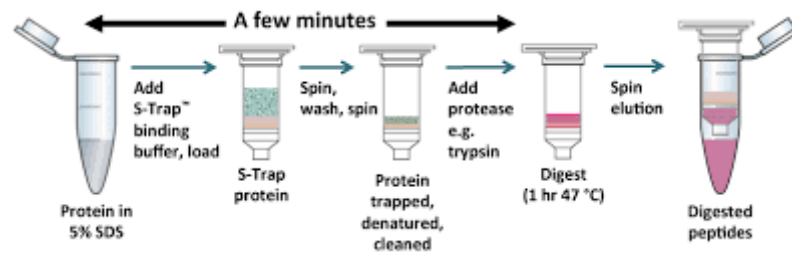
- C18 reverse-phase resin
- Wash buffer: 0.5% ACN, 0.1% formic acid
- Elution buffer: 70% ACN, 0.1% formic acid



# Higher-Throughput Sample Prep Options



Thermo EasyPep



Protifi S-Trap